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The Effect of the Combination of Vitamin K₂ and Genistein, Coumestrol and Daidzein on the Osteoblast Differentiation and Bone Matrix Formation

تاثير التداخل لفيتامين K₂ مع الجنستين، الكومسترول والديادزين عل تمايز خلايا الارومة العظمية وتكوين المادة العظمية

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Abstract

Multiple studies have been reported the stimulatory effect of the combinations of nutrients factors on bone formation. One such factor is vitamin K_2 which can be associated with bone protective activities. The effect of vitamin K_2 alone and in combination with genistein, coumestrol and daidzein on osteoblast differentiation and mineralization were tested. Significantly, vitamin K_2 increased bone mineralization in combination with genistein (10^{-5} M), coumestrol (10^{-7} M) and daidzein (10^{-5} M). However, there is no additive effect of this vitamin on alkaline phosphatase (ALP) levels in osteoblasts. By contrast, vitamin K_2 enhanced the stimulatory effect of type I collagen and osteocalcin expression. Vitamin K_2 alone increased RUNX and OSX expression while there is no synergistic effect with tested compound; this vitamin also did not modulate nuclear factor kappa B ligand (RANKL)/ osteoprotegerin (OPG) ratio expression. These results suggested that vitamin K_2 can be more effective factor in the presence of phytoestrogens on the improvement of bone formation after menopause.

Keywords: vitamin K2, phytoestrogens, osteoblast, bone resorption, bone matrix

الملخص

يعتبر فيتامين K₂ احد العناصر المهمة التي تتصاحب مع حماية العظم. تمت في هذه الدراسة تقييم تاثير هذا الفيتامين لوحده او بوجود المركبات الفلافونيدية الجنستين ، الكومسترول والديادزين على فعالية الخلايا البادئة للعظم وترسيب المعادن . بينت الدراسة ان فيتامين K₂ بوجود المركبات الفلافونيدية قد حفزت معنويا ترسيب المعادن في العظم والتي تزيد من صلابته بينما لم يوجد اي تاثير على فعالية انزيم Alkaline المركبات الفلافونيدية قد حفزت معنويا ترسيب المعادن في العظم والتي تزيد من صلابته بينما لم يوجد اي تاثير على فعالية انزيم Phosphatase المركبات الفلافونيدية قد حفزت معنويا ترسيب المعادن في العظم والتي تزيد من صلابته بينما لم يوجد اي تاثير على فعالية انزيم Phosphatase Phosphatase ايضا فان هذا الفيتامين مع المركبات الفلافونيدية قد حفز التعبير الجيني للجينات التي لها دور في بناء البروتينات العظمية الكولاجين والكالسين العظمي بينما لم توجد اي زيادة في ه ذا التعبير بوجود المركبات الفلافونيدية . كما لم يوجد تأثير على التعبير الجيني الكولاجين والكالسين عن مستقبلات التي لها دور في تمايز الخلايا الناقضة للعظم وعن جينات تمايز الخلايا العظمية . للجينات التي تعبر عن مستقبلات التي لها دور في تمايز الخلايا الناقضة للعظم وعن جينات تمايز الخلايا البادة للعظم . وينت هذه الدراسة ان فيتامين K₂ نمو داخلي البادئة للعظم .</sub> بينت هذه الدراسة ان التراسة ان للجينات التي تعبر عن مستقبلات التي لها دور في تمايز الخلايا الناقضة للعظم وعن جينات تمايز الخلايا البادئة العظم .

الكلمات الدالة: فيتامين k₂، فيتوستروجين، بناء العظم، ارتشاف العظم، مصفوفة العظم

Introduction

The skeleton remodels in response to changes in mechanical load, Ca^{2+} and damage [1,2]. Bone remodeling is performed by osteoblasts that secrete and mineralize new bone matrix and osteoclasts that resorb bone. Osteoblast and osteoclast are regulates during bone formation and any disruption causes bone resorption such as post-menopausal osteoporosis and osteomyelitis [3]. Previous study shown that phytoestrogens (PEs) suppressed osteoclast differentiation and bone resorption, induced osteoblast differentiation and bone matrix formation as well [4] and also increased bone matrix formation following phytoestrogen supplementation in women [5,6].

Vitamin K identified as a fat-soluble nutrient required for coagulation and also acts as a co-factor for carboxylase, an essential enzyme for γ -carboxylation of vitamin K-dependent osteocalcin and Gla proteins [7]. However, the main function of vitamin K is the maintenance of the blood clotting proteins levels that synthesized as inactive proteins in the liver. Importantly, vitamin K species (K₁ and K₂) is known to promote γ -carboxylation of glutamic acid residues to vit K-dependent osteocalcin bone matrix and also possess both anti-osteoclastic and pro-osteoblastic action [8,9]. Moreover, vitamin K₂ has long been considered as potent protective factor against osteoporosis, hepatocarcinoma, and atherosclerosis [10,11]. Interestingly, several clinical trials detected the positive correlation between bone fracture and vitamin K deficiency [12-14].

Additionally, vitamin K_1 acts as antioxidant agent, whereas vitamin K_2 (MK-4) proposed as a protective agent for neural cells from apoptosis by decreasing oxidative stress [15]. Flavones found to contain soybeans, menaquinon-7 (K_2) and cryptoxanthin, which have been revealed to stimulate osteoblastic bone formation and inhibit osteoclatic bone resorption *in vitro*. It was found that the supplementation with vitamin K_2 prevented bone loss in ovariectomised (OVX) rats, and its intake has a stimulatory effect on bone mass in human. Therefore, accumulated studies paid the attention toward the important role of vitamin K in bone metabolism.

Natural vitamin K_2 (MK-7) with seven isoprene unites is abundant in fermented soybeans (*natto*) and has anabolic effect on bone calcification. In contrast, vitamin K_2 has been caused a significant increase in alkaline phosphatase (ALP) activity and calcium content in femoral-metaphyseal tissues *in vitro*. Furthermore, MK-7 (10⁻⁶ or 10⁻⁵M) caused a significant increasing in calcium content, ALP activity and DNA contents in cortical bone and trabecular bone in ovx rats. The oral administration of Vitamin K_2 to neuroectomised rats also was found to reduce trabecular bone loss, prevention of osteoblast dysfunction, and increasing bone formation rate [16]. In addition, this effect enhanced in combination with genistein (10⁻⁵, 10⁻⁶M), suggestion the mode of action of this vitamin is differs from genistein, and MK-7's activity was abolished in the presence of cyclohexamide [17]. Thus, the combination of these two factors may be useful in the prevention of bone resorption and disorders. Recent study found that vitamin K increased the mineralisation in primary human cells, although this was associated with decreasing in type-I collagen expression [18]. Vitamin K_2 also influenced bone formation of Fas expression on osteoblast treated with or without TNF- α [19].

Several studies suggest that the dietary factors such as Zn and phytoestrogens have a positive effect on bone cell activity [4, 20- 22]. However, these studies have not fully examined the effect of combinations of phytoestrogens (PEs) and Vit K_2 on osteoblast differentiation and activity. Therefore, the current study examined the effect of genistein, coumestrol and daidzein in the presence of vitamin K_2 on osteoblast differentiation, function and bone matrix formation *in-vitro*.

Materials and Methods

Cell culture

Saos-2 human osteoblast like cells were obtained from ECACC, Porton Down, UK and cultured in RPMI1640 media supplemented with 10% charcoal stripped fetal calf serum, 2mmol/l glutamine, 100IU/mL benzylpenicillin and 100 mg/mL streptomycin [23].

Effect of Vitamin K₂ on alakaline phosphatase (ALP) activity and mineralization

The effect of coumestrol, daidzein and genistein at $(10^{-5} \text{ to } 10^{-9} \text{ M})$ concentration, in the presence or absence of vitamin K₂ (10^{-5} M) on alkaline phosphatase activity was assessed as follows. Saos-2 cells were incubated incubated with treatments in presence of β -glycerophosphate (β -GP) (10 mM) and L-ascorbic acid (L-AA) (50 mg/l) for four days. ALP activity measured by staining cultures with *p*-nitrophenyl phosphate (1 mg/ml) at 37°C for 30 min [24]. Absorbance was measured at 405 nm. Mineralization was assessed using a modification of Hale's methodology [25]. After 15 days of incubation culture, the cells incubated with medium containing 1 μ g/ml calcein for four hours at 37°C, and fluorescence measured by a cytofluor II fluorescence multiwell plate reader (Perseptive Biosystem, USA).

Real time quantitative PCR analysis

Saos-2 cells were incubated with PEs with or without vitamin K₂. Total RNA was extracted using a Sigma genelute RNA isolation kit and reverse-transcribed with Moloney Murine Leukemia Virus RT using random nonamer primers. Real-time PCR was performed on a StepOne PCR system (Applied Biosystems) using the DNA-binding dye SYBR green for detection of PCR products. Primers for genes were as follows: human Runtrelated transcription factor 2 (Runx2) forward AGACCCCAGGCAGGCACAGT, reverse GCGCCTAGGCACATCGGTGA; human osterix forward GCACCCTGGAGGCAACTGGC, reverse GAGCTGGGTAGGGGGCTGGA; human type I collagen forward CCTGGCAGCCCTGGTCCTGA, reverse CTTGCCGGGCTCTCCAGCAG; human receptor activator of NF-kB ligand [26] forward ACAGGCCTTTCAAGGAGCTGTGC, reverse ACCAGATGGGATGTCGGTGGC; human osteoprotegerein (*OPG*) forward AATCGCACCCACAACCGCGT reverse AGCAGGAGACCAAAGACACTGCA; human β actin forward GCGCGGCTACAGCTTCACCA, reverse TGGCCGTCAGGCAGCTCGTA.

Statistical analysis

Differences between groups were assessed using Fisher's one-way ANOVA (SPSS Inc., Chicago, IL, USA). A difference of $P \le 0.05$ was considered statistically significant.

Results and Discussion

Effect of vitamin K₂ on RANKL/OPG ratio and Mineralization

This study found that vitamin K_2 alone or with genestein and coumestrol had no direct effect on RANKL/OPG ratio expression, increased the inhibitory effect of daidzein on this ratio, however this was not significant when compared to control or daidzein alone Figure (1).



Fig. (1): The effect of vitamin K₂ on the expression of nuclear factor kappa B ligand (RANKL)/osteoprotegerin (OPG) Ratio RANKL/OPG ratio). G10⁻⁷= genistein 10⁻⁷M, C10⁻⁷= coumestrol 10⁻⁷M, D10⁻⁵= daidzein 10⁻⁵M.

(Vitamin K_2 alone had no significant effect on mineralization; the addition of vitamin K_2 significantly enhanced the mineralization induced by genistein (10⁻⁵ M) Figure (2A), coumestrol (10⁻⁷M) Figure (2B), and daidzein (10⁻⁵M, 10⁻⁶M) Figure (2C) in comparison to test compounds alone , indicating the positive effect of vitamin K_2 on inorganic bone minerals deposition.



Fig. (2): Synergistic effect of vitamin K₂ with phytoestrogens on bone mineralization. * Values are significantly different from phytoestrogens treatment alone. Gen= genistein, Cou= coumestrol, Dai= daidzein.

Effect of vitamin K₂ on ALP activity

The results found that vitamin K_2 had no effect on ALP activity in combination with coumestrol or genistein, although these compounds alone and in combination with vitamin K_2 increased ALP expression versus control group Figure (3A, B). However, the addition of vitamin K_2 enhanced ALP expression with daidzein (10⁻⁵ M) Figure (3C).



Fig. (3): Synergistic effect of of vitamin K₂ with phytoestrogens on ALP activity. * P≤0.05 versus phytoestrogens treatment alone. Gen= genistein, Cou= coumestrol, Dai= daidzein.

Effect of vitamin K₂ on type I collagen, osteocalcin, Runx2 and osx gene expression

Vitamin K_2 alone decreased *type I collagen* or *osteocalcin* expression, but vitamin K_2 addition augmented the stimulatory effect of coumestrol and daidzein on *type I collagen* and *osteocalcin* expression and had no effect appeared in combination with genistein on *type I collagen* expression and also reduced the stimulatory effect of genistein on *osteocalcin* expression to the comparable level of control group Table (1). Vitamin K_2 alone increased *Runx2* and *osterix* expression; however this vitamin did not enhance the stimulatory effect of genistein, coumestrol and daidzein on both gens expression. However, All tested compounds have been shown to induce *Runx2 and Osterix* expression versus control group, coumestrol (10⁻⁷ M), daidzein (10⁻⁵ M) and genistein (10⁻⁷ M) Table (2).

Table (1): The effect of vitamin K ₂ on the expression	organic components of bone matrix in combination with
genistein, coumestrol and daidzein	

	Type I collagen mRNA copies		Osteocalcin mRNA copies	
	Mean	SE	Mean	SE
Control	154110.8	12003.	770	78.1
Vitamin K ₂ (10 ⁻⁵ M)	70212.8	5452.9	441 ^a	36.3
Coumestrol (10 ⁻⁷ M)	1069198 ^a	75038	1868.4 ^a	217.5
Coumestrol (10 ⁻⁷ M) and K ₂	1398595.7 ^{a b}	122951	2892.8 ^{a b}	386.6
Daidzein (10 ⁻⁵ M)	62457.5 ^a	3934.9	1190.1 ^a	263.4
Daidzein (10 ⁻⁵ M) and K ₂	438989.8 ^{a b}	43731.2	1865.1 ^{a b}	181.5
Genistein (10 ⁻⁷ M)	97275.6 ^a	3368.2	1607 ^a	209
Genistein (10 ⁻⁷ M) and K ₂	67680 ^b	7037.4	685 ^{a b}	62

a P≤0.05 versus control, b P≤0.05 versus relevant PEs alone.

	Runx2 mRNA copies		Osterix mRNA copi	ies
_	Mean	SE	Mean	SE
Control	1521.3	295	27355	6643.9
Vitamin K_2 (10 ⁻⁵ M)	22538 ^a	1025.6	69965.7 ^a	18836.3
Coumestrol (10 ⁻⁷ M)	28582 ^a	858.8	139140.7 ^a	10415.5
Coumestrol (10 ⁻⁷ M) and K ₂	25911 ^a	480.6	124928 ^{ab}	9519.6
Daidzein (10 ⁻⁵ M)	99419 ^a	7357.9	138813.2 ^a	25524.5
Daidzein (10 ⁻⁵ M) and K ₂	103024 ^{ab}	11717	147171.3 ^{ab}	9677.3
Genistein (10 ⁻⁷ M)	27610 ^a	975.9	115361.2 ^a	10062.9
Genistein (10 ⁻⁷ M) and K ₂	31192 ^a	1091	128211.2 ^{ab}	8281.4

Table (2): The effect of vitamin K₂ on RUNX2 and osterix expression in combination with genistein, coumestrol and daidzein

a P≤0.05 versus control group.

Bone remodeling is a process including bone resorption and bone formation that generates a skeleton optimized to mechanical and mineral requirements. Several studies observed the positive effect of dietary factors such as phytoestrogens on bone cell differentiation and activity. However the cellular mechanism mediating the action of the combination of PEs and vitamin K₂ on osteoblasts is still open area to debate. This study clarifies the cellular mechanism through the appropriate combination of these factors may augment osteoblast differentiation. Bone resorption is regulated by osteoblast signals that stimulate osteoclast formation from monocytic precursors. Calcium fall in serum act a resorptive stimuli that increase RANKL expression and decreased in OPG (soluble decoy receptor for RANKL) expression on osteoblast surface [27]. Vitamin K₂ has long been considered as potent protective factor against osteoporosis, hepatocarcinoma, and atherosclerosis [10, 11]. Interestingly, several clinical trials detected the positive correlation between bone fracture and vitamin K deficiency [12-14].

In this study, Vitamin K_2 significantly increased the mineralization peak induced by genistein, coumestrol and daidzein concluded that vitamin K_2 has protective effect on non-organic matrix bone formation. The stimulatory effect of vitamin K_2 on mineralization was noted also on organic bone matric, *type I collagen* and *osteocalcin* expression in combination with phytoestrogens, indicating that the vitamin K_2 can enhance the anabolic effect of phytoestrogen in appropriate concentration and increased bone matrix formation.

In opposite, vitamin K_2 has not been shown to enhance ALP activity induced with phytoestrogens except with daidzein at (10⁻⁵ M) concentration which this activity significantly increase after vitamin K_2 addition, although ALP activity still significant versus control group. The present result suggested that the augmentative effect of vitamin K_2 on mineralization did not mediate through ALP activity.

This study aimed to examine the mechanism by which phytoestrogens and zinc or vitamin K_2 augment osteoblast function, current study determined the expression of key regulators of osteoblast differentiation. Osteoblastogenesis is a sequential process involving transcription factors that lead to stimulate mesenchymal precursors to form mature osteoblasts [28]. The initial stage of osteoblast differentiation is controlled by the expression Runx2 and Osterix.

Vitamin K_2 alone increased *Runx2* and *Osx* expression but there is no additive effect of this vitamin on the anabolic effect of anti-osteoclastic dose of genistein, coumestrol and daidzein on early key expression responsible for oseoblastic expression RANKL/OPG ratio) This results is consistent with ALP activity that explained there is no effect on early stage on osteoblast differentiation while the major effect was detected on bone matrix formation, type I collagen and osteocalcin expression which is consistent with mineralization results, which may be the mechanism mediated the positive effect of this vitamin on bone formation.

In addition to the effect of vitamin K_2 on RUNX2 and Osx, it is also increased the peak of osteocalcin and type I collagen levels. Additionally, vitamin K_2 was noticed directly induced both organic and non-organic bone formation and no effect appeared on early stage and ALP activity of osteoblast differentiation and osteoclastic stimuli RANKL and OPG expression which may be the mechanism mediated the anabolic effect of this vitamin on bone formation.

These findings conducted that vitamin K_2 can be more effective in combination with tested phytoestrogens through different mechanisms involving increasing of osteoblasts differentiation and mineralization. The current study found that the anabolic effect of vitamin K_2 mediated in biphasic way, vitamin K_2 can enhance mineralization, type I collagen and osteocalsin formation, an organic bone matrix, and no effect on RANKL/OPG ratio. Thus, appropriate combination of these two factors may alter bone formation as found *in vivo* [29, 30].

These results strengthen the data for the use of combinations of PEs with vitamin K_2 in the treatment of skeletal disorders and these dietary factors can improve bone formation after menopause through different mechanisms. **References**

- 1. Martin, T. J.and Seeman, E. (2008). Bone remodeling: its local regulation and the emergence of bone fragility. Best Practice & amp; Research Clinical Endocrinology & amp. Metabolism. 22: 701-722.
- Hnriksen, K., Neutzsky-Wulff, A.V., Bonewald, L.F. and Karsdal, M.A. (2009). Local communication on and within bone controls bone remodeling. Bone. 44: 1026-1033.
- 3. Manolagas, S.C., Kousteni, S. and Jilka, R.L. (2002). Sex Steroids and Bone. Recent Progress in Hormone Research. 57: 385-409.
- **4.** Karieb, S. and Fox, S.W. (2011). Phytoestrogens directly inhibit TNF-α-induced bone resorption in RAW264.7 cells by suppressing c-fos-induced NFATc1 expression. Journal of Cellular Biochemistry. 112: 476-487.
- Mei, J., Yeung, S. S. C. and Kung, A.W.C. (2001). High Dietary Phytoestrogen Intake Is Associated with Higher Bone Mineral Density in Postmenopausal but Not Premenopausal Women. The Journal of Clinical Endocrinology and Metabolism. 86: 5217-5221.
- 6. Weaver, C. M., Martin, B. R., Jackson, G. S., Mccabe, G. P., Nolan, J. R., Mccabe, L. D., Barnes, S., Reinwald, S., Boris, M. E.and Peacock, M. (2009). Antiresorptive Effects of Phytoestrogen Supplements Compared with Estradiol or Risedronate in Postmenopausal Women Using 41Ca Methodology. Journal of Clinical Endocrinology and Metabolism. 94: 3798-3805.
- 7. Berkner, K. L. (2000). The Vitamin K-Dependent Carboxylase. The Journal of Nutrition. 130: 1877-1880.
- **8.** Yamaguchi, M. and Ma, Z. J. (2001). Inhibitory effect of menaquinone-7 (vitamin K₂) on osteoclast-like cell formation and osteoclastic bone resorption in rat bone tissues in vitro. Molecular and Cellular Biochemistry. 228: 39-47.
- **9.** Yamaguchi, M., Uchiyama, S. and Tsukamoto, Y. (2002). Stimulatory effect of menaquinone-7 on bone formation in elderly female rat femoral tissues in vitro: prevention of bone deterioration with aging. International Journal of Molecular Medicine. 10(6):729-33.
- 10.Vermeer, C., Shearer, M.J., Zittermann, A., Bolton-Smith, C., Szulc, P., Hodges, S., Walter, P., Rambeck, W., Stocklin, E. and Weber, P. (2004). Beyond Deficiency: Potential benefits of increased intakes of vitamin K for bone and vascular health. European Journal of Nutrition. 43: 325-335.
- **11.** Iwamoto, J., Takeda, T. and Sato, Y. (2004). Effects of vitamin K₂ on osteoporosis. Current Pharmaceutical Design., 10(21):2557-76.
- 12. Booth, S. L., Broe, K. E., Gagnon, D. R., Tucker, K. L., Hannan, M. T., Mclean, R. R., Dawson-Hughes, B., Wilson, P. W. F., Cupples, L. A. and Kiel, D. P. (2003). Vitamin K intake and bone mineral density in women and men. The American Journal of Clinical Nutrition. 77: 512-516.
- 13. Feskanich, D., Weber, P., Willett, W. C., Rockett, H., Booth, S. L. and Colditz, G. A. (1999). Vitamin K intake and hip fractures in women: a prospective study. The American Journal of Clinical Nutrition. 69: 74-79.
- 14. Ebina, K., Noguchi, T., Hirao, M., Kaneshiro, S., Tsukamoto, Y. and Yoshikawa, H. (2015). Comparison of the effects of 12 months of monthly minodronate monotherapy and monthly minodronate combination therapy with vitamin K₂ or eldecalcitol in patients with primary osteoporosis. J. Bone Miner Metab.
- **15.** Li, J., Wang, H. and Rosenberg, P. A. (2009). Vitamin K prevents oxidative cell death by inhibiting activation of 12-lipoxygenase in developing oligodendrocytes. Journal of Neuroscience Research. 87: 1997-2005.
- 16. Iwasaki, Y., Yamato, H., Murayama, H., Takahashi, T., Ezawa, I., Kurokawa, K. and Fukagawa, M. (2002). Menatetrenone prevents osteoblast dysfunction in unilateral sciatic neurectomized rats. The Japnanese Journal of Pharmacology. 90 (Hirano): 88-93.
- **17.**Yamaguchi, M., Uchiyama, S. and Tsukamoto, Y. (2002). Stimulatory effect of menaquinone-7 on bone formation in elderly female rat femoral tissues *in vitro*: prevention of bone deterioration with aging. International Journal of Molecular Medicine. 10 (6): 729-733.
- **18.** Atkins, G. J., Welldon, K. J., Wijenayaka, A. R., Bonewald, L. F. and Findlay, D. M. (2009). Vitamin K promotes mineralization, osteoblast-to-osteocyte transition, and an anticatabolic phenotype by γ -carboxylation-dependent and -independent mechanisms. American Journal of Physiology Cell Physiology. 297: C1358-C1367.
- 19. Urayama, S., Kawakami, A., Nakashima, T., Tsuboi, M., Yamasaki, S., Hida, A., Ichinose, Y., Nakamura, H., Ejima, E., Aoyagi, T., Nakamura, T., Migita, K., Kawabe, Y. and Eguchi, K. (2000). Effect of vitamin K₂ on osteoblast apoptosis: Vitamin K₂ inhibits apoptotic cell death of human osteoblasts induced by Fas, proteasome inhibitor, etoposide, and staurosporine. Journal of Laboratory and Clinical Medicine. 136: 181-193.
- **20.**Poulsen, R. C. and Kruger, M. C. (2008). Soy phytoestrogens: impact on postmenopausal bone loss and mechanisms of action. Nutrition Reviews. 66: 359-374.
- **21.**Seo, H.J., Cho, Y.E., Kim, T., Shin, H.I. and Kwun, I.S. (2010). Zinc may increase bone formation through stimulating cell proliferation, alkaline phosphatase activity and collagen synthesis in osteoblastic MC3T3-E1 cells. Nutrition Research and Practice. 4(5): 356-361.

- **22.**Bhardwaj, P., Rai, D.V and Garg, M.L. (2013). Zinc as a nutritional approach to bone loss prevention in an ovariectomized rat model. Menopause. 20 (11):1184-93.
- **23.** Gallagher, J.A. (2003). Human osteoblast culture, in Bone Research Protocols (Helfrich, M.H. and Ralson, S.H., eds.) Humana, Totowa, NJ.
- 24. Sabokbar, A., Millett, P. J., Myer, B. and Rushton, N. (1994). A rapid, quantitative assay for measuring alkaline phosphatase activity in osteoblastic cells *in vitro*. Bone Miner. 27: 57-67.
- **25.** Hale, L.V., MA, Y.F. and Santertte, R.F. (2000). Semi-quantitative fluorescence analysis of calcein binding as a measurement of *in-vitro* mineralization. Calcified Tissue International. 67: 80-84.
- 26. Tang, Z., Sahu, S. N., Khadeer, M. A., Bai, G., Franklin, R. B. and Gupta, A. (2006). Overexpression of the ZIP1 zinc transporter induces an osteogenic phenotype in mesenchymal stem cells. Bone. 38: 181-198.
- **27.**Boyce, B. F. and Xing, L. (2008). Functions of RANKL/RANK/OPG in bone modeling and remodeling. Archives of Biochemistry and Biophysics. 473: 139-146.
- 28.Jensen, E. D., Gopalakrishnan, R. and Westendorf, J.J. (2010). Regulation of gene expression in osteoblasts. Bio Factors. 36: 25-32.
- **29.** Ehara, Y., Takahashi, H., Hanahisa, Y. and Yamaguchi, M. (1996). Effect of vitamin K₂ (menaquinone-7) on bone metabolism in the femoral-metaphyseal tissues of normal and skeletal-unloaded rats: enhancement with zinc. Research in Experimental Medicine (Berl). 196(3): 171-178.
- **30.**Pearson, D.A. (2007). Bone health and osteoporosis: the role of vitamin K and potential antagonism by anticoagulants. Nutrition in Clinncal Practice., 22(5):517-44.