The effect of infection with *Entamoeba histolytica* on the level of some biological variables and histological changes in the liver

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ABSTRACT

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Background: Entamoeba histolytica is an intestinal protozoan parasite that causes "dysentery," leading to Amebic colitis and a liver abscess. Infection begins with ingesting infective stages, represented by the cystic form found in contaminated food and drink; the parasite attacks the tissues by attaching to the epithelial lining of the intestine, and adhesion occurs through virulence factors. **Objective:** Detection of the E. histolytica parasite using the direct swab method, study the histological changes in the liver and some blood parameters of animals infected with E. histolytica and compare these with uninfected groups. Material and method: The current study was conducted between May 1, 2021, and October 30, 2022, to diagnose infection with E. histolytica. Samples were taken from clinically infected patients who suffered from diarrhea and examined microscopically using a direct wet swab. In the experimental study, E. histolytica was given to laboratory mice. Male laboratory mice were divided into four groups. The first group represented the negative control group, dosed with Normal saline only. Results: The negative control group showed normal liver histological sections. The hepatic lobule appeared to include hepatic cells, which were arranged radially, as they appeared as cords extending from the central vein, in addition to diagnosing normal liver cells. With central round nuclei and a homogeneous appearance of cytoplasm, hepatic lamellae, and hepatic sinusoids, the second group represented the positive control group. It was treated with the parasite E. histolytica, which recorded numerous histological changes in the livers of the group, which included irregular radiological appearance of the hepatic cells around the hepatic vein, infiltration of lymphocytes, and necrosis and swelling of some hepatic cells, there was an increase in liver enzymes, indicating infection with E. histolytica. Conclusion: Laboratory animals infected with E. histolytica had histological changes in the liver, represented by necrosis, nucleolytic, and amoebic liver abscesses. Although the parasite infects the intestines and settles there, it causes secondary infections through its transmission through the blood and lymph to the liver.

Keywords: Histological changes, liver, enzymes, E.histolytica. DOI: https://doi.org/10.24126/jobrc.2024.18.2.786

1-INTRODUCTION

The disease caused by the parasite *Entamoeba histolytica* is called intestinal amoebiasis (colitis). Ulceration begins in the intestine in the rectum and appendix area or at the top of the colon and may spread along the length of the colon. The increase in the number of parasites in the ulceration area increases the destruction of the mucous layer. The ulcer is at the basement membrane or musculoskeletal layer and then begins to erode gradually, causing a superficial area of coagulation to occur. This initial ulcer is accompanied by bacterial invasion, and the cellular response in the host is minimal (1).

In chronic infections, the amoeba invades the mucous layer, decomposes the submucosal layer, and penetrates the muscular and serous layers. Thus, the nutrient phase can be transmitted through the blood and lymph to other organs, hence forming secondary infections. A high percentage of deaths occur as a result of colon penetration with peritonitis. Sometimes, a granular mass called an amoeboma forms in the intestinal wall, which leads to intestinal obstruction. This is due to the host's cellular response to chronic ulcers, which usually contain active nutrient phases. Secondary infections can be found in all body organs, but the liver is the most common organ. It is most affected in about 5% of cases (2).

2-Material and Method

Parasite Diagnosis of macroscopic Examination:

This method included describing the shape and texture of stool samples as soft, light, watery, or semisolid, indicating their type of parasitic organisms. The vegetative stages of intestinal primary animals are often found in a soft, light sample, while the cystic stages of these parasites appear in large, hard samples (3). It was also observed whether the stool sample was bloody or mucous, as the presence of blood and mucus indicates an infection, so samples containing blood and mucus are given special care. In such a case, the sample must be taken from the blood-stained and mucus-containing areas. It should also be noted whether the stool is greasy; the sample's color and smell, which may be moldy or spoiled, should be noted.

Laboratory animal groups

Examining the feces of the mice before starting the experiment to ensure that they were free from infection with parasitic intestinal infections by placing a small number of feces on a glass slide and mixing it with a bit of Logul's Iodine, the slide was covered and examined under a microscope, (10) and then mice were killed as they were considered as a negative control mice group. Then, ten mice were dosed with amoebic sludge after the dose was determined, as it was given orally using Gavage. The animal was dosed by holding it in a way that did not allow it to move. Gavage was introduced into the mouth, and after making sure that it was inserted into the esophagus, the dose was slowly pushed into the stomach of the animal. After the dose Mice infected with *Entamoeba histolytica* were placed in clean cages, and stool was collected and examined by microscope to determine the incidence of infection. The examination took place 48 hours after the dose, and after confirming the presence of infection, a group of mice was taken, and stool samples were collected from them and kept in a tube. They were considered a positive control group and were not treated with any treatment. Testing Blood samples

Blood biochemical parameters were determined using a 1.5 ml blood sample collected by cardiac puncture, transferred to blood collection tubes, and centrifuged at $3,000 \times g$ for 15 minutes. Serum was collected to determine aspartate aminotransferase (A.S.T) and alanine aminotransferase (A.L.T) by using RANDOX kits to estimate the effectiveness of the GOT and GPT enzymes in serum respectively as recommended by the manufacturing company according to the method of (4). The ALP (Alkaline phosphatase) enzyme level was estimated according to the method mentioned in the ALP determination kit, according to the method of (5).

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3-Results

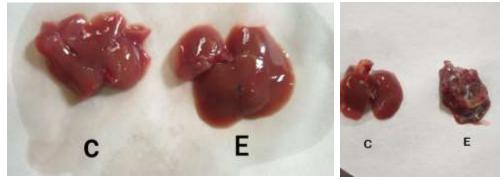
Table (1) shows the Variations in the levels of AST, ALT, and ALP in animals infected with *E. histolytica* infection:

			Enzymes
ALP I.U/L	(ALT) GPT I.U/L	(AST) GOT I.U/L	Group
a 24.38 ± 2.47	31.2±1.61 a	32.75±4.79 a	Negative control
b 46.25 ± 4.95	47.67±4.23 b	57.44±2.87 b	Positive control

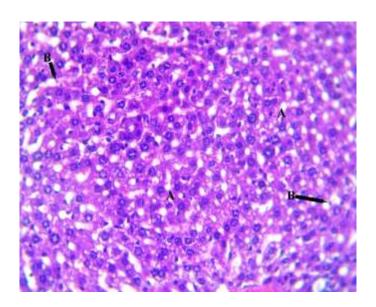
*Different letters indicate significant differences (probability ≤ 0.05) between the rates of the different groups.

E. histology Morphological changes of the liver

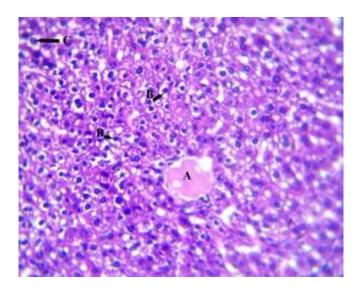
The results of the visual examination of mice infected with the parasite *E. histolytica* showed congestion and enlargement of the liver. Liver deformity reached a large degree in the infected groups compared to control groups in which the liver appeared normal and without deformities. Congestion was observed in the external appearance of the liver, with a change in color and the appearance of dark areas. The liver showed blood congestion and dark scarring.



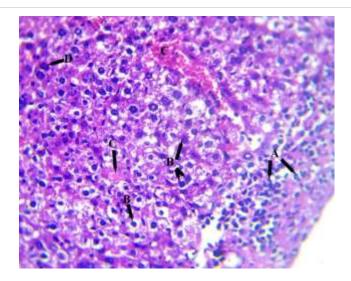
The letter C indicates the liver of the negative control group, while the letter E indicates the positive control group infected with *E. histolytica*.



Figure(1): A section of the liver tissue of the negative control group showing (A) the liver tissue containing rows of tangled liver cells, (B) the blood sinusoids containing red blood cells and Kupffer cells (H&E 400X).



Figure(2): Liver tissue of the positive control group infected with *E. histolytica*, showing (A) a central vein with decomposed blood, (B) hyperplasia of liver cells with bulging of the cytoplasm and thickening of their nuclei, (C) parasitic particles (H&E 400X).



Figure(3): Liver tissue of the positive control group infected with *E.histolytica*, showing (A) lymph node infiltration in the liver tissue, (B) hyperplasia of liver cells with cytoplasmic bulging and thickening of the nuclei, (C) blood congestion in the sinusoids, (D)) Parasitic particles (H&E 400X).

4-Discussion

The results of the current study recorded an increase in the concentration of aminotransferase enzymes (A.L.T, A.S.T) in groups infected with *E. histolytica* compared to the negative control group, as in Table (1). In general, the A.L.T enzyme is considered specialized for liver cell activity. In contrast, the A.S.T. enzyme is usually found in various tissues of the body, especially the heart, liver, kidney, and skeletal muscle. In general, any damage that occurs in liver cells due to the invasion of the parasite, which possesses some tissue-degrading enzymes, leads to the release of the enzyme into the blood, thus increasing its concentration. This explains the increased effectiveness of (A.L.T and A.S.T) for groups infected with *E. histolytica*; Liver alkaline phosphatase (ALP) levels and acute liver abscesses have increased AST and ALT levels, Aminotransferase levels are sensitive indicators of liver cell injury and help identify liver cell diseases like liver abscess; therefore, these enzyme levels serve as markers. More specifically, when the liver cell membrane is broken, both enzymes are released into the bloodstream at greater levels. Liver cell senescence also confirms the release of aminotransferases (6).

The ALP enzyme showed highly significant differences in the second group (Positive control) compared to the First group (Negative control), However, the differences did not reach significance between all groups, and the initial group was also not noteworthy in contrast to the control group. These enzymes are typically found in muscle cells to a lesser degree and mostly in liver cells. Aspartate aminotransferase (AST) and alanine aminotransferase (ALT) levels in the blood rise when the liver is injured or damaged because liver cells leak these enzymes into the bloodstream. ALP is a material in the bile ducts of the bones, intestines, and liver; it signifies liver disease. High levels of ALP can result from bile duct damage or obstruction, and these tests can reveal a variety of details regarding a range of pathological alterations (6). This is in line with the findings of (7), who noted that in patients with diarrhea, there was an increase in the level of AST and ALT enzymes in the serum, as 90% of cases had an increase in the ALP level, this outcome agreed with the findings of (8) because the serum levels of liver enzymes in patients with parasite infection are incredibly high, and (9) showed a rise in liver enzyme levels. However, this is at odds with the data that have been collected. (10) That patients' liver function tests showed elevated levels. The current study agreed with the study of (11), which indicated that the survey revealed that in addition to altering the biochemical parameters (ALP, AST, and ALT), the *E. histolytica* parasite also caused histological changes in the organs, particularly the liver, such as programmed cell death.

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E. histology Morphological changes of the liver showed that the mucous layer of the intestine, with its hydrolytic enzymes, causes its perforation and is transmitted to other organs, such as the liver, causing abscesses. This is consistent with the results of (12) and (13), who indicated the occurrence of macroscopic changes and congestion in the internal organs of infected mice with *E. histolytica*.

An amoebic liver abscess (ALA) occurs most commonly after *E. histolytica* colonizes the large intestine, causing intestinal dysentery. In about 1% of cases, the parasite damages the intestinal mucosa and spreads to other organs, leading to various dysentery symptoms beyond the Intestines (14) and (15).

The destruction of host tissues and the survival of amoebae in the liver is associated with a robust adaptive response and regulation of proteins, including amoeboid virulence factors. (16).

The adhesion molecule Gal/Gal NAC lectin, cysteine proteins, and other virulence factors of *E. histolytica* have received the most research. (17), protein amoeba (18), and Liposomes phosphoglycans (19). Because of their significant contribution to ALA etiology in humans and animals.

Collagen, fibrinogen, elastin, and laminin are components of extracellular materials that the parasite must penetrate to cause invasive disease. *E. histolytic*'s cysteine proteins can break down these materials (15). These proteins also play a role in breaking down cellular monolayers (20). It has also been suggested that cysteine proteins (CP) contribute to creating the anaerobic environment that trophozoites require for growth in vivo during ALA development (21) and (22).

They cause eukaryotic cells and bacteria to lyse (18), have a proven cytolytic ability, and are implicated in inducing necrosis and apoptosis in vivo (23).

An amoebic liver abscess (ALA) formation following intraportal inoculation with *E. histolytica* consists of three successive stages: acute inflammation, abscess formation, and necrosis. Microabscess formation and hepatocyte damage have occurred, culminating in tissue necrosis a week after infection (24), demonstrating that distant hepatocytes die from necrosis and that amoebic particle diffusion takes place in the endothelium. These authors speculate that the secretion of amoebic particles, which have the ability to produce harmful effects at a distance, may be the cause of cytotoxicity, even in the absence of direct trophozoite-hepatocyte contact. This study found that the number of apoptotic cells increased with the length of incubation. It had fragmented nuclei, which are important characteristics of apoptosis. Indicating a gradual increase in apoptosis in infected slices during the increasing incubation period, the development of ALA causes severe liver tissue destruction, consistent with a study (15).

The appearance of abundant infiltration of inflammatory cells and the occurrence of bleeding and congestion are due to structural changes in the liver tissue that lead to the secretion of chemoattractive factors and then the infiltration of inflammatory cells, such as monocytes, to defend the body; Inflammation is part of a biological response. It is a complex process through which the body tries to get rid of pathogens and other factors to begin the healing process; when any foreign body or toxic materials enter the living body, the inflammatory process begins with the first step by identifying the foreign substance, and then blood vessels begin to expand, and fluids and inflammatory cells filter into nearby tissues, where they are attracted to the site of injury to treat inflammation (25).

The parasite's toxins affect the liver's metabolic activity, as metabolic activity increases when the liver is exposed to toxic substances resulting from the metabolism of pathogens and through the process of detoxification for the purpose of balancing Oxidative stress resulting from the action of toxins, and this increase is in order to release energy sources such as glucose, which is accompanied by cell death, decomposition, and necrosis, which occurs after severe degeneration or occurs directly (26).

5-Conclusion

Laboratory animals infected with *E. histolytica* had histological changes in the liver, represented by necrosis, nucleolytic, and amoebic liver abscesses. Although the parasite infects the intestines and settles there, it causes secondary infections through its transmission through the blood and lymph to the liver.

Reference

- 1- Steve CS, Chadee K., *Entamoeba histolytica*: Host parasite interactions at the colonic epithelium. Tissue Barriers, (2017); 5(1): 1-4.
- 2- Roberts L.S., Janovy J. Foundation of parasitology. 7thed. McGraw-Hill companies, London, (2005): 108-111.
- 3- Kudo RR. Protozoology, 5th edn., Charles C. Thomas publ., Parasitol., (1966); 101: 322-326.
- 4- Singh GR, Manjappara UV. Impromptu Effects of Nt8U with Soyasaponins on Obesity-related Lipid Parameters in High Fat Fed C57BL/6 Mice. International Journal of Peptide Research and Therapeutics. (2020); 26(1): 405-411.
- 5- Iorhemba AP, Imolele AG. The effect of combined treatment of vitamin C and loperamide on intestinal sodium and potassium ion ATPase, alkaline phosphatase and lipid peroxidation on castor oil induced diarrheal rats. bioRxiv. (2020).
- **6-** Dufour, D. Evaluation of liver function and injury in clinical (Henry J editor). W. B. Saunders Company, (2001). 264 pp.
- 7- Al-Kubaissi, Abdul Wahab Badawi Hussien. Immunological and Epidemiological Study of Patients Infected with *Entamoeba histolytica*. Ph.D. thesis, College of Science, AL – Mustansiriya University, (2002) 125 pp.
- Pluta, H., Pluta, J.N. Hepatic abscess : current approach to patients with pyogenic or amebic abscess. Gastroenterologia polska., (2008); 15(5): 343-346.
- 9- Al Ghanimi, Fatima Yusuf Ktan. Study the opposite effect for some plant extracts on the parasite Giardia lamblia in mice infected laboratory. Master Thesis, College of Science, University ofAl-Muthanna., (2013); 130 pp.
- 10- Fernandes, H.; Souza, C.R.; Swethadri, G.K., Naik, C.R. Ameboma of the colon with amebic liver abscess mimicking metastatic colon cancer. Indian Journal of Pathology and Microbiology. (2009); 52(2): 228-230.
- 11- Chabuk; Halla Abdul-Hadi Abdul-Ghani, Al-Saadi ;Haidar Kamil Zaidan, Al-Hamairy; Ahmed Khudhair Obayes. Biochemical Changes of Liver That Infected With *Entamoeba histolytica* In White Rats, Journal of Babylon University/Pure and Applied Sciences, (2014); No.(9),Vol.(22).
- 12- Jabak, Hala Abdel Hadi Abdel Ghani. Study of some histological and functional changes in male rats infected with *Amoeba histolytica*. Master Thesis. College of Science for Girls. University of Babylon. (2013).
- 13- Al-Rafi'i, Amal Kamel Abdel-Sada Jabr. Isolation and diagnosis of *Entamoeba histolytica* in humans and study of some pathological and biochemical changes in mice experimentally infected with the parasite. Master's thesis. College of Veterinary Medicine, University of Baghdad. (2014).
- 14- Stanley, S.L. Amoebiasis. Lancet. (2003); 36: 481-489.
- 15- Carranza-Rosales, P.; Santiago-Mauricio, M.G.; Guzman-Delgado, N.; Vargas- Villarreal, J.; Lozano-Garza, G.; Viveros-Valdez, E.; Ortiz-Lopez, R.; Moran- Martinez, J. and Gandolfi, A.J. Induction of virulence factors, apoptosis, and cytokines in precision-cut hamster liver slices infected with *Entamoeba histolytica* Experimental Parasitology,(2012); 132:424-433.
- 16- Bruchhaus, I., Roeder, T., Lotter, H., Schwerdtfeger, M., Tannich, E., Differential gene expression in *Entamoeba histolytica* isolated from amoebic liver abscess. Molecular Microbiology, (2002); 44: 1063-1072.
- 17- Bruchhaus, I.;Loftus, B. ;Hall, N., Tannich ,E. The intestinal protozoan parasite *Entamoeba histolytica* contains 20 cysteine proteins genes, of which only a small subset is expressed during in vitro cultivation. *Eukaryot Cell*. (2003); 2(30): 501- 509. doi:10.1128/EC.2.3.501-509.

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- 18- Leippe, M.; Bruhn, H.; Hecht, O., Grotzinger, J. Ancient weapons: the three dimensional structure of amoebapore A. Trends in Parasitology, (2005); 21: 5-7.
- 19- Moody-Haupt, S., Patterson, J.H., Mirelman, D., McConville, M.J. Themajor surface antigens of *Entamoeba histolytica* trophozoites are GPI-anchored proteophosphoglycans. Journal of Molecular Biology. (2000); 297: 4-8.
- 20- Lauwaet, T.; Oliveira, M.J.; Callewaert, B.; De Bruyne, G.; Mareel, M. and Leroy, A. Proteinase inhibitors TPCK and TLCK prevent *Entamoeba histolytica* disturbance of tight junctions and microvilli in enteric cell layer in vitrol. International Journal of Parasitology, (2004); 34: 785-794.
- 21- Pérez -Tamayo, R.; Montfort, I.; Olivos Garcia, A.; Ramos, E.; Nequiz, M. and Tello, E,. Amibiasis hepatica. Revista de Gastroenterologia de México., (2006); 71: 47–72.
- 22- Zhang, X.; Zhang, Z.; Alexander, D.; Bracha, R.; Mirelman, D., Stanley, S.L. Expression of amoebapores is required for full expression of *Entamoeba histolytica* virulence in amebic liver abscess but is not necessary for the induction of inflammation or tissue damage in amebic colitis. Infection and Immunity, (2004); 72: 678-683.
- 23- Andra, J.; Herbst, R., Leippe, M, Amoebapore archaic effector peptides of protozoan origin are discharged into phagosomes and kill bacteria by permeabilizing their membranes. Developmental & Comparative Immunology., (2003); 27: 291–304.
- 24- Ventura-Juarez, J.; Campos-RodrIguez, R.and Tsutsumi, V. Early interactions of *Entamoeba histolytica* trophozoites with parenchymal and inflammatory cells in the hamster liver: an immunocytochemical study. Canadian Journal of Microbiology., (2002); 48: 123–131.
- **25-** Johnson L. V., Anderson D. H., Mullins R. F., Hageman G. S. A role for local inflammation in the formation of drusen in the aging eye. Am. J. Ophthalmol. (2002); 134(3): 411-431.
- 26- Murray R. K., Granner D. K. Mayes P. and Rodwell V. W. Harpers illustrated biochemistry 26th ed. Alange. Med. (2003).

تأثير الإصابة بـ Entamoeba histolytica على مستوى بعض المتغيرات البيولوجية والتغيرات النسجية في الكبد

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خلفية البحث: يعد طفيلي Intestinal protozoan parasites ، ويؤدي إلى حدوث خراج الكيد Intestinal protozoan ، ويؤدي إلى حدوث خراج الكيد Liver abscess ، نيدأ العدوى بابتلاع الأدوار المعدية Amoebiasis والمتمثلة بالشكل المتكيس Colitis amoeba ، ويؤدي إلى حدوث خراج الكيد يعيجم الطفيلي الانسجة عن بابتلاع الأدوار المعدية Amoebiasis والمتمثلة بالشكل المتكيس colitis amoeba ، ويؤدي إلى حدوث خراج الكيد Liver abscess على بابتلاع الأدوار المعدية infective stags والمتمثلة بالشكل المتكيس costic form الموجود في الشراب والطعام الملوث ويهاجم الطفيلي الانسجة عن طبيلي الأدوار المعدية infective stags ، ويحدث الالتصاق بواسطة عوامل ضراوة virulence factors ، **هدف البحث** : الكشف عن طفيلي *طر*يق التصاقه بالبطانة الظهارية للمعي ، ويحدث الالتصاق بواسطة عوامل ضراوة virulence factors ، **هدف البحث** : الكشف عن طفيلي ومقارنتها مع المجاميع غير المصابه الم**واد وطريقة العمل** : أحريت الدارسة الحالية خلال الفترة ما بين آليار الدموية للحيوانات المصابه بعرض عرض ومقارنتها مع مالمعابير الدموية للحيوانات المصابه بعد *E.histolytica* ومقارنتها مع المجاميع غير المصابه ال**مواد وطريقة العمل** : أحريت الدارسة الحالية خلال الفترة ما بين آليار 2021 و 203 ولاريقا للمعادي وعريقة العمل : أحريت الدراسة الحالية خلال الفترة ما بين آليار 2021 و لاتشرين الأول 2022 بغرض ومقارنتها مع المجاميع غير المصابه ال**مواد وطريقة العمل** : أحريت الدار مى المصابون سريرياً يعانون من الإسهال عندما تم فحص العينات مجهريا باستخدام المحرية وينا المالقرة ، وفي الدراسة التجريبية ، تم تقسيم ذكور فئران المختبر إلى مجموعتين منالمجموعة الاولى مجموعة السيطرة السيطرة السابح وي سريرياً يعانون من الإسهيالي عندما تم فعلي عليما تصويلي المولى مجموعة المالمراقي نوى ميتنا مع مرون المالي عموم عنا و ألى العريفة تحريعها فمويا بطفيلي الحال النسيج : علمموعة الولى مجموعة البيال لحد وأن الفصيص المجري يشمل خلايا كبية ومظهر متبعن للمالي على العامي الحال للنبي عمون من الوريد المحموعة الثانية مترون المحيوم على توى سيس التعيور المالي بالغيلي الحمو ع الثانية متلت ماميو وإلى المحيوية وي مسيص العبي ألى معاون فرى مولى مرون وألى معرون وألى وألى معاول فرض مروى وألى فلال علي المي وي مالمموع عن الولي لعرم من المولى عروى مالم محر

الكلمات المفتاحية: تغيرات نسجية ، كبد ، انزيمات ، الاميبا الحالة للنسيج.